

U.S. Serial No. 10/555,073

Amendment in response to Notice to Comply filed January 21, 2011

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**Amendments to the Specification:**

Please amend the specification as shown:

Please insert the following heading and paragraph on page 1, below the title:

**Sequence Listing**

**The instant application contains a Sequence Listing which has been submitted in ASCII format via EFS-Web and is hereby incorporated by reference in its entirety. Said ASCII copy, created on January 18, 2010, is named 308410US.txt and is 462,420 bytes in size.**

Please amend the paragraph on page 19, lines 29-30 as follows:

**Figure 6** shows the amino acid sequence of the SARS virus M (Matrix) Glycoprotein (residues 1-220 of SEQ ID NO: 34).

Please amend the paragraph on page 20, lines 7-12 as follows:

**Figure 10A-B** show an alignment of the nucleocapsid protein N from the SARS virus (Tor2\_N; SEQ ID NO: 36) and various other nucleocapsid proteins (SEQ ID NOs: 44-52; and SEQ ID NO: 199 of AIBV2 nucleocapsid protein [Avian infectious bronchitis virus 2]). Asterisks (\*) indicate percentage identity to the SARS nucleocapsid protein calculated by Align (Myers and Miller, CABIOS (1989) 4:11-17). **Figures 11-A-K** show the nucleotide sequence of the 29,751-base genome of the SARS virus (SEQ ID NO: 15).

Please amend the paragraph on page 22, lines 17-20 as follows:

**Figure 27** shows an alignment of the secreted region of the SARS virus ORF 10 (SEQ ID NO: 201) of the 29,751-base genome sequence (sars) with the conotoxin from *Conus ventricosus* (conotoxin) (SEQ ID NO: 200). Sequence identity is indicated by asterisks and sequence homology is indicated by dots.

Please amend the paragraph on page 60, lines 18-27 as follows:

More specifically, size-selected cDNAs were ligated into the pCR4-TOPO TA cloning vector (Invitrogen, CA), or after digestion with the restriction nuclease Not I into the pBR194c vector (The Institute for Genomic Research, Rockville, MD, USA). Ligated clones were then transformed by electroporation into DH10B T1 cells (Invitrogen), plated on 22 cm agar plates with the appropriate

antibiotic and grown for 16 hours at 37°C. Colonies were picked into 384-well Axygen culture blocks containing 2 X YT media and grown in a shaking incubator for 18 hours at 37°C. Cells were lysed and DNA purified using standard laboratory procedures. Sequencing primers for the 194c clones were 5'-GGCCTCTTCGCTATTACGC-3' (forward primer) **(SEQ ID NO: 159)** and 5'-TGCAGGTCGACTCTAGAGGAT-3' (reverse primer) **(SEQ ID NO: 198)**.